

FIG. 1

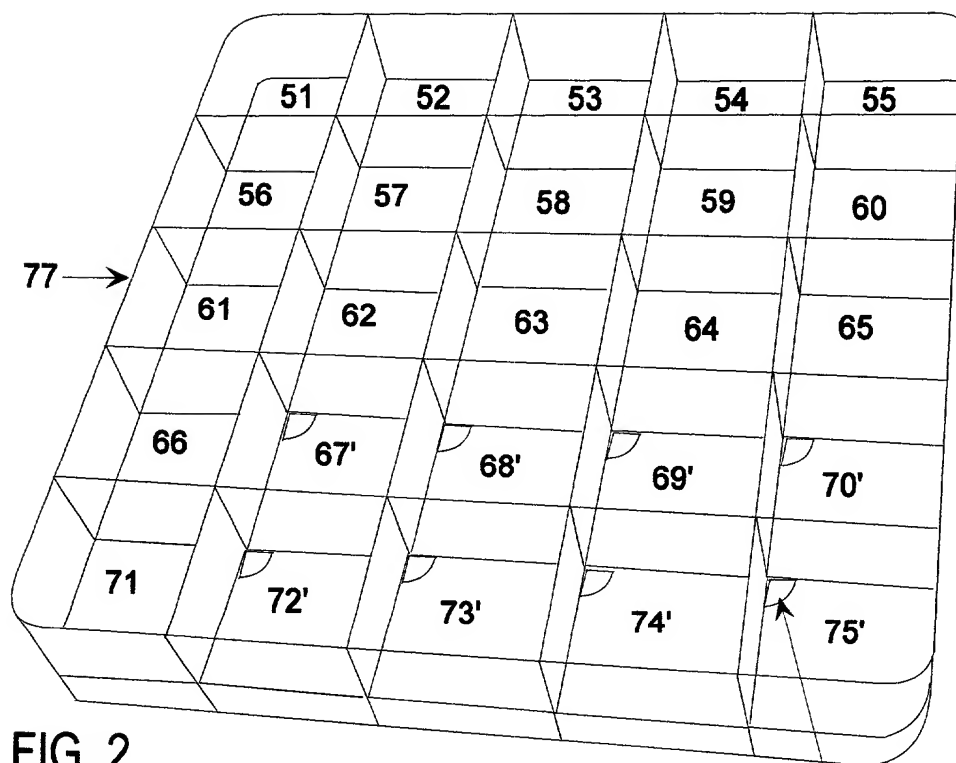


FIG. 2

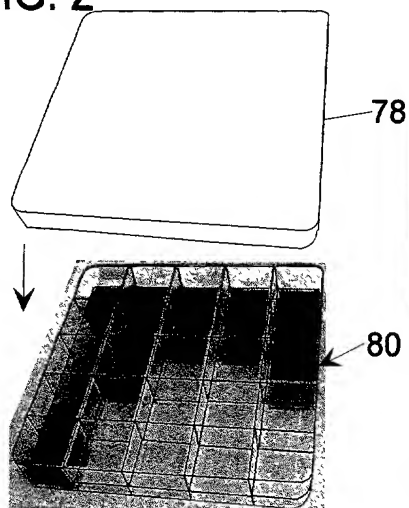


FIG. 3

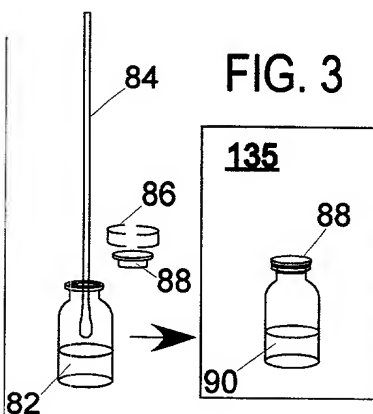


FIG. 4

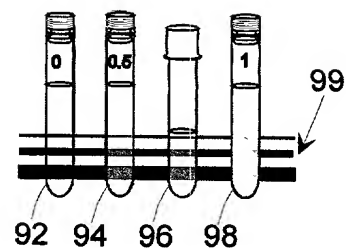


FIG. 5

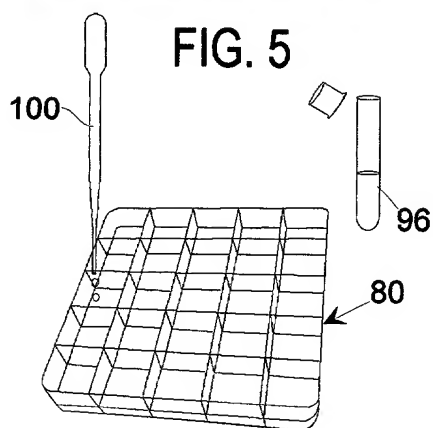


FIG. 6

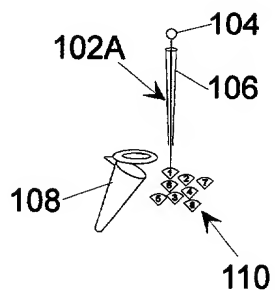


FIG. 37A

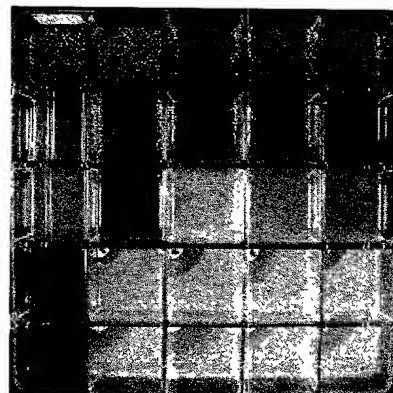


FIG. 7
Prior Art

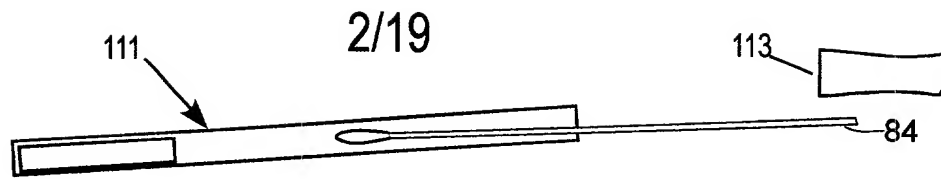


FIG. 8A
Prior Art

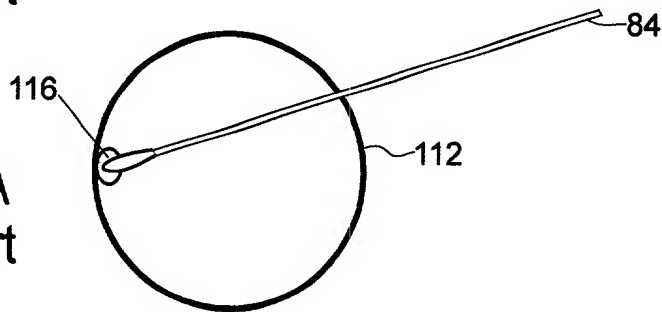


FIG. 8B
Prior Art

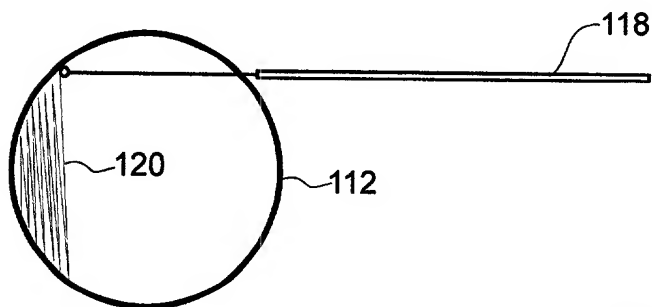


FIG. 8C
Prior Art

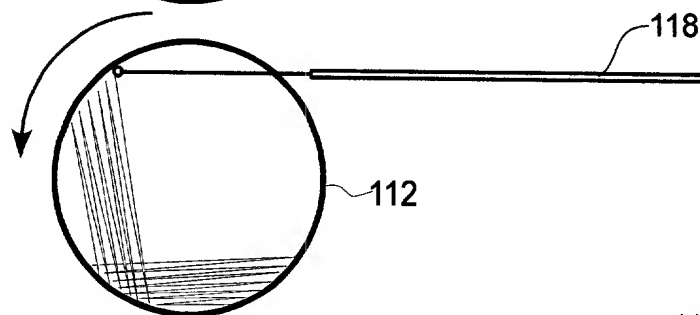


FIG. 8D
Prior Art

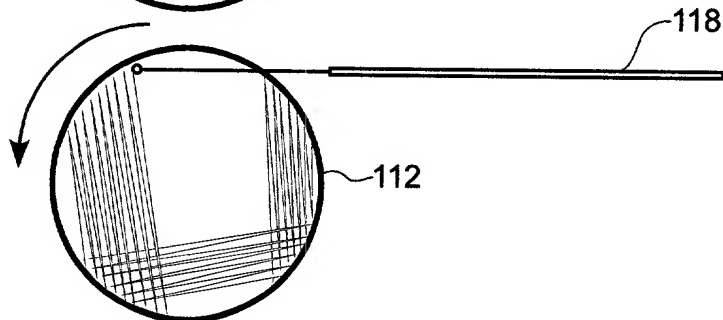


FIG. 8E
Prior Art

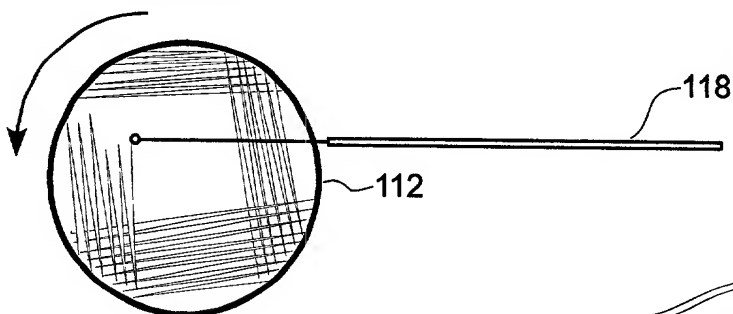


FIG. 9
Prior Art

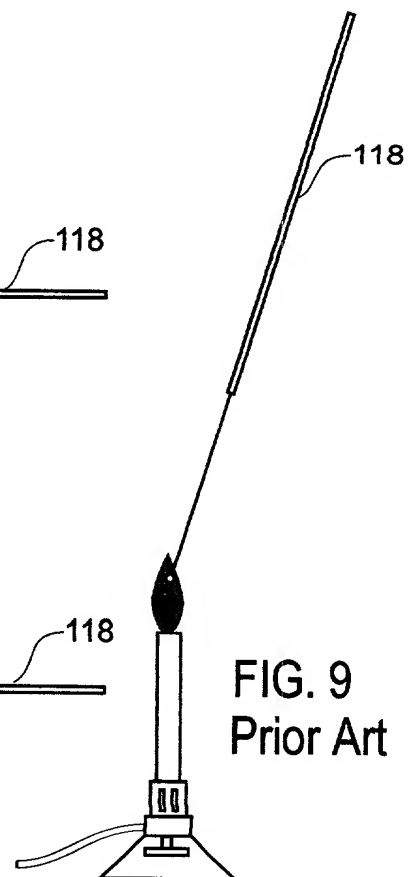
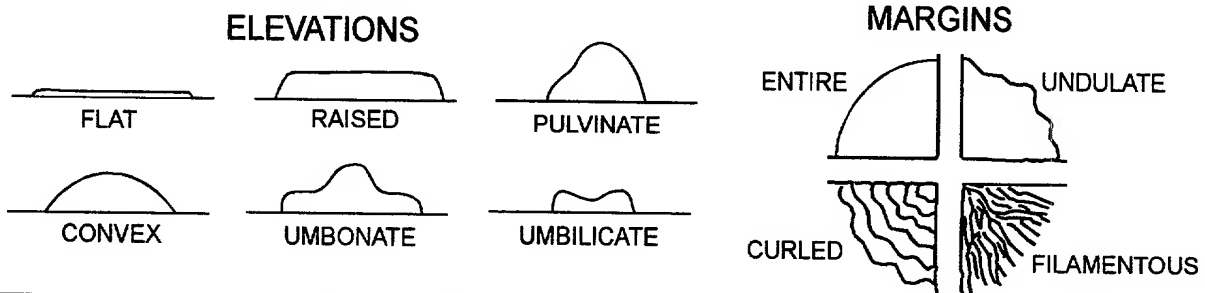














TABLE OF COLONY CHARACTERISTICS



FORM: OVERALL SHAPE OF COLONY WHEN VIEWED FROM TOP	<p>CIRCULAR - MOSTLY ROUND, MAY BE SLIGHTLY UNEVEN</p> <p>WRINKLED - UNEVEN SURFACE TEXTURE, OFTEN DRY IN APPEARANCE</p> <p>IRREGULAR - EDGES VERY UNEVEN</p> <p>RHIZOIDAL - BRANCHED (UNCOMMON)</p> <p>FILAMENTOUS - POWDERY, SPREADING LINES (FUNGI)</p> <p>CURLED - SEPARATED EDGES, CONCENTRIC CIRCLES (UNCOMMON)</p>
ELEVATION: VIEW COLONY FROM SIDE	<p>FLAT - WHEN LIGHT IS REFLECTED ACROSS SURFACE OF COLONY, NO CONVEX SHAPE IS SEEN (COMMON)</p> <p>RAISED - ELEVATED (COMMON)</p> <p>CONVEX - SLIGHT DOME SHAPE (COMMON)</p> <p>PULVINATE - HAT-LIKE APPEARANCE -DOMED IN MIDDLE, SLIGHTLY RAISED AT EDGES (MOST SIGNIFICANT IN 1-2 DAY CULTURE)</p>
MARGIN: VIEW EDGE OF COLONY	<p>ENTIRE - SMOOTHLY CURVING EDGE (COMMON)</p> <p>UNDULATE - WAVY EDGE (COMMON)</p> <p>LOBATE - VERY IRREGULAR AMOEBA-LIKE EDGES (UNCOMMON)</p> <p>FILAMENTOUS - POWDERY LINES (FUNGI)</p> <p>CURLED - SEPARATE EDGES (UNCOMMON)</p>
CONSISTENCY: TEXTURE OF COLONY WHEN LOOP IS INSERTED INTO IT	<p>BUTYROUS - BUTTER-LIKE, CAN PICK UP PASTE EASILY</p> <p>MUCOID - SLIMY (<3MM OF SLIME THAT ATTACHES TO END OF LOOP WHEN SAMPLING) (COMMON)</p> <p>VISCID- STICKY, RESISTANT TO PICK UP OF PASTE, SOME ELASTICITY OF COLONY HAS BEEN LOST (UNCOMMON)</p> <p>WAXLIKE - COLONY FRAGMENTS WHEN BEING PICKED UP (UNCOMMON)</p> <p>POWDERY - LIGHT FILAMENTS (FOUND MOSTLY IN MOLDS)</p>
PIGMENT (COLOR)	<p>OFF-WHITE - ANY VARIATION ON WHITE, INCLUDING GREY, TAN, CREAM, IVORY VERY PALE YELLOW</p> <p>WHITE - PORCELAIN WHITE ONLY (UNCOMMON)</p>
APPEARANCE (REFLECTED LIGHT)	<p>TEXTURE AS YOU TILT PLATE AND VIEW AT AN ANGLE FROM THE TOP UNDER A BRIGHT LIGHT</p>
APPEARANCE (TRANSMITTED LIGHT) VIEW WHILE HOLDING UP TO BRIGHT LIGHT AND LOOK THROUGH COLONIES.	<p>TRANSPARENT - COMPLETELY SEE-THROUGH, COLONIES HARD TO SEE (UNCOMMON).</p> <p>TRANSLUCENT - CAN SEE MODEST REDUCTION OF LIGHT PASSING THROUGH COLONY (COMMON)</p> <p>OPAQUE - ALMOST NO LIGHT PASSES THROUGH COLONY (COMMON)</p>
DIAMETER OF COLONY	<p>COLONIES NEED TO BE WELL SEPARATED. FUNCTION OF TIME</p>

FIG. 11
Prior Art

TABLE OF CELLULAR MORPHOLOGY BY LIGHT MICROSCOPE OBSERVATION (400X-600X). MOTILITY WILL BE OBSERVED WITH SOME OF THESE.		
 <p>Spherical cells (COCCI)</p>	 <p>Rod-shaped cells (BACILLI)</p>	 <p>Spiral-shaped cells (SPIRILLA)</p>
 <p>Streptococci</p>	 <p>Coccobacilli (short rods)</p>	 <p>Borrelia-type spirillum</p>
 <p>Staphylococci</p>	 <p>Coryneform bacilli (club shaped rods)</p>	 <p>Treponema-type spirillum</p>
 <p>Diplococci</p>	 <p>Vibrio (comma shaped rods)</p>	 <p>Leptospira-type spirillum</p>

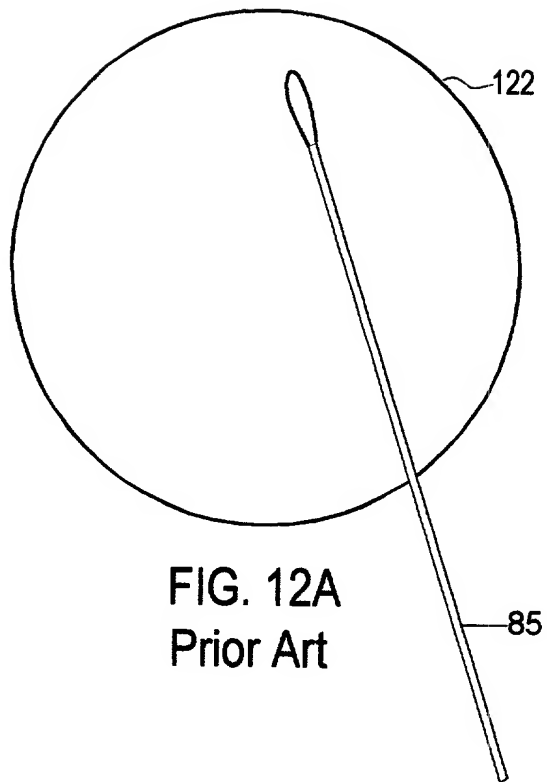


FIG. 12A
Prior Art

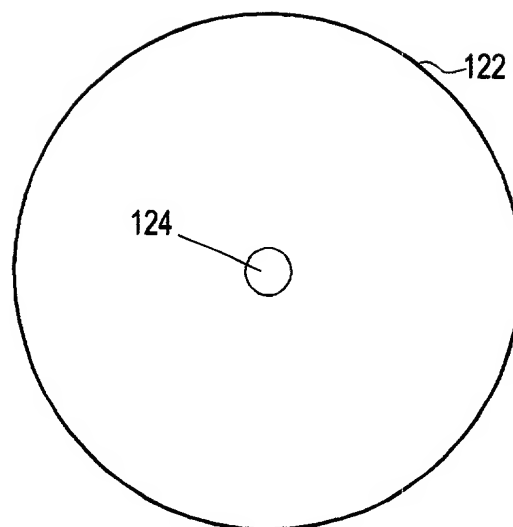


FIG. 12B
Prior Art

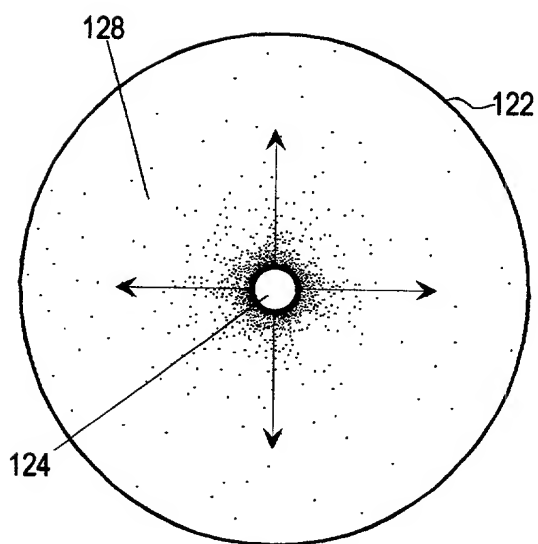


FIG. 12C
Prior Art

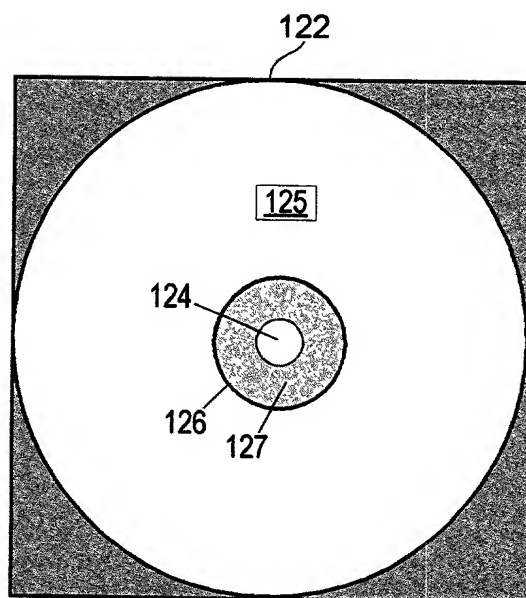


FIG. 12D
Prior Art



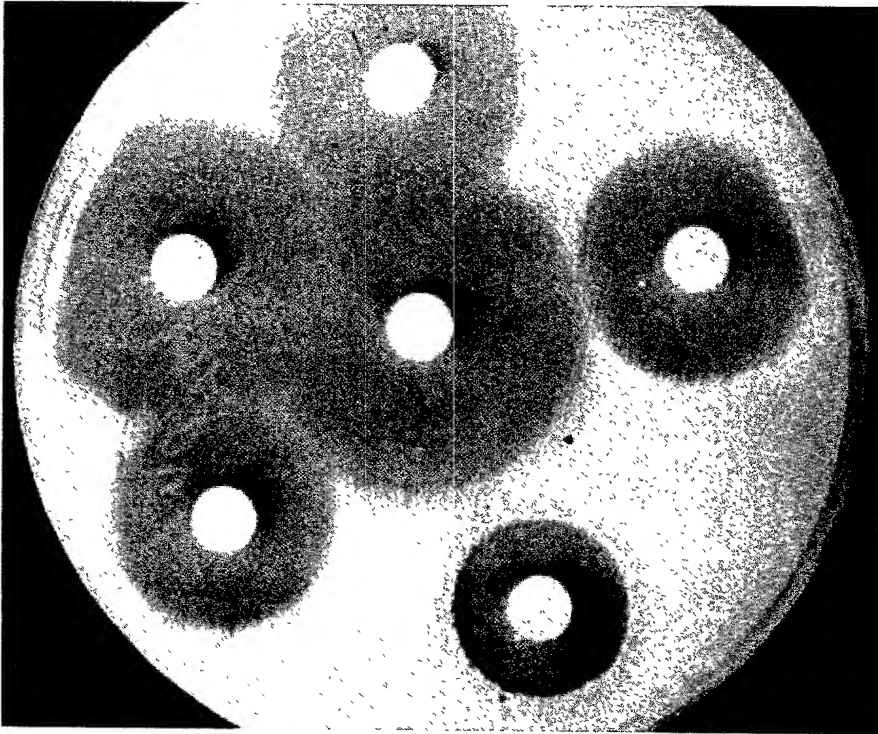


FIG.13
Prior Art

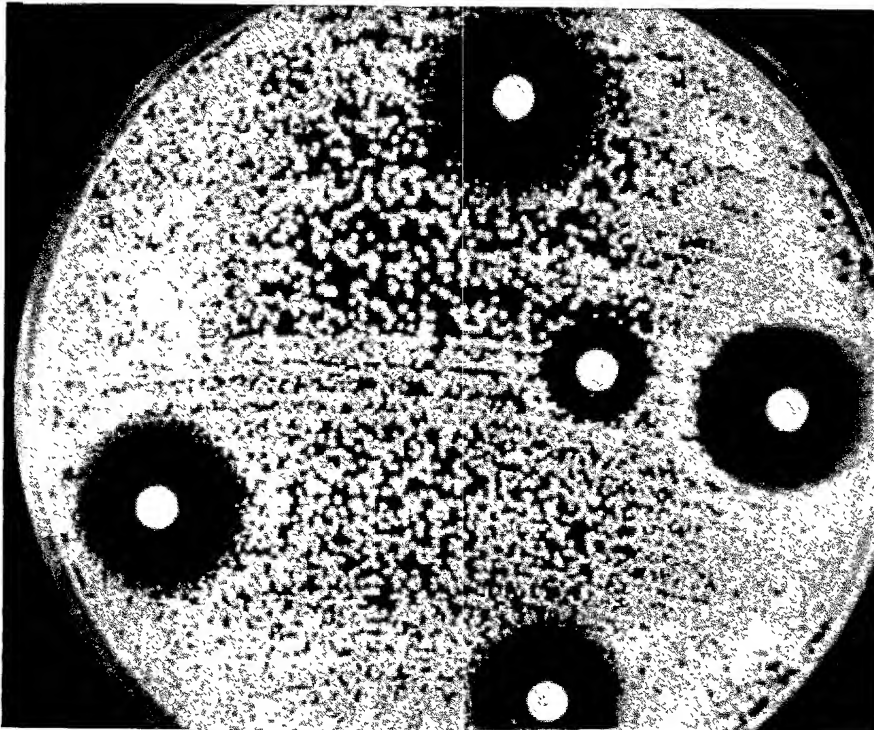


FIG.14
Prior Art

FIG. 13



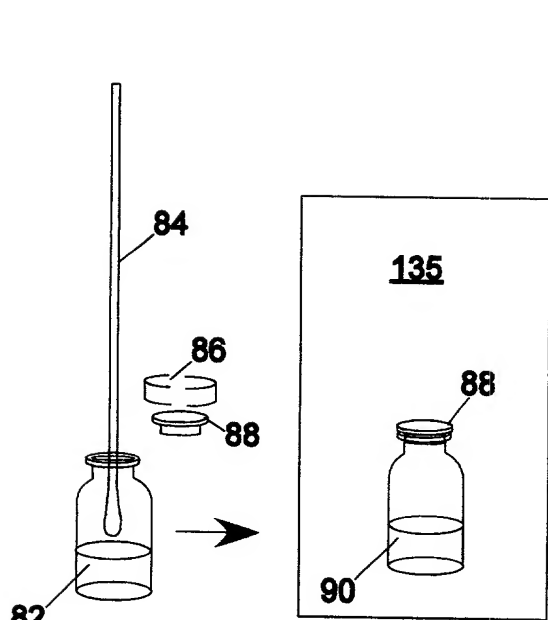


FIG. 3

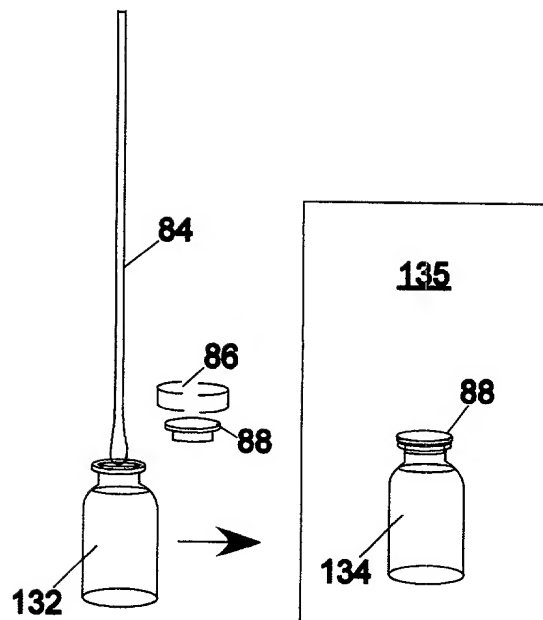


FIG. 15

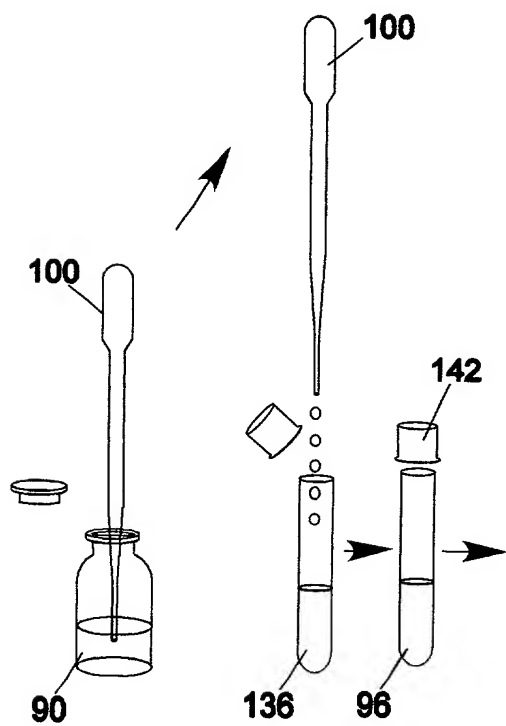


FIG. 16A

FIG. 16B

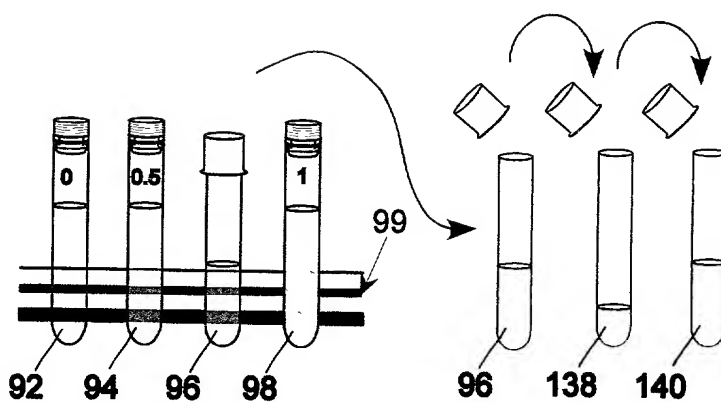


FIG. 16C

FIG. 16D



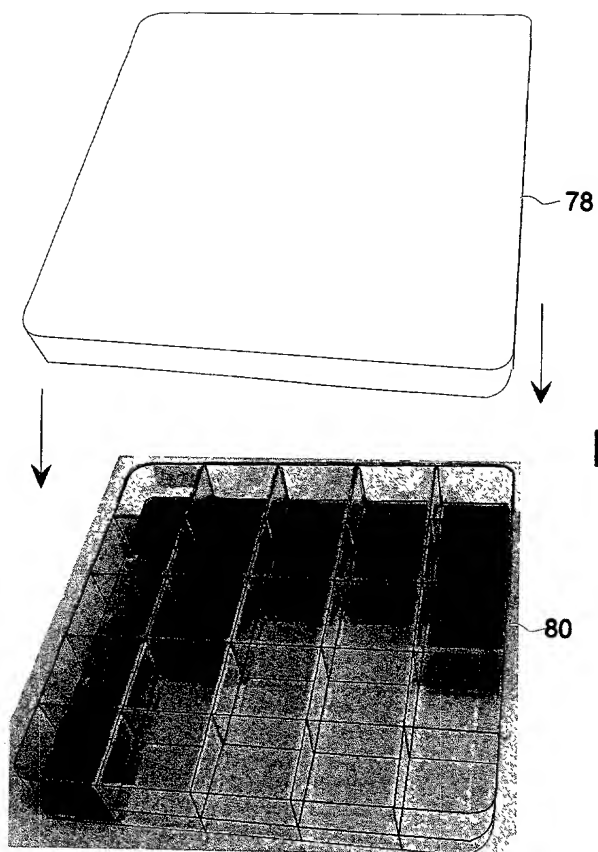


FIG. 2

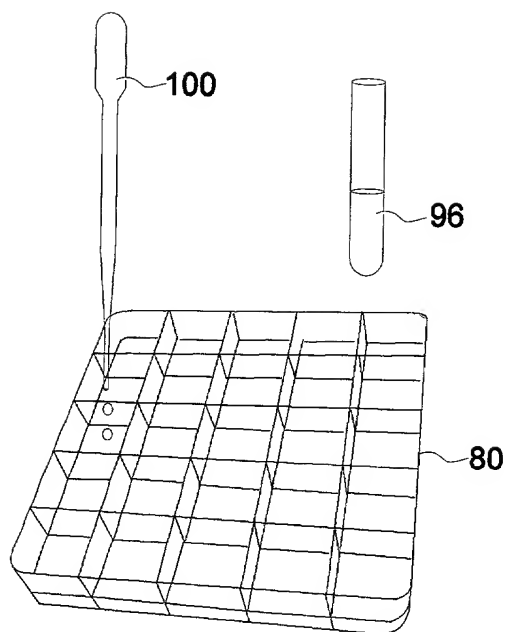


FIG. 5

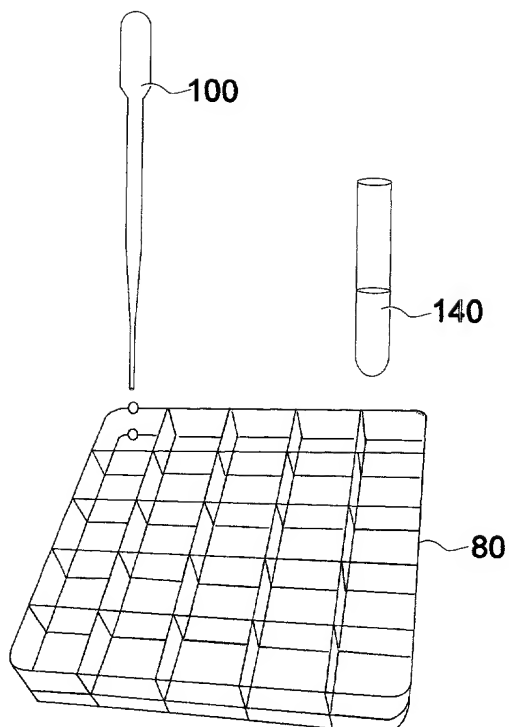


FIG. 17



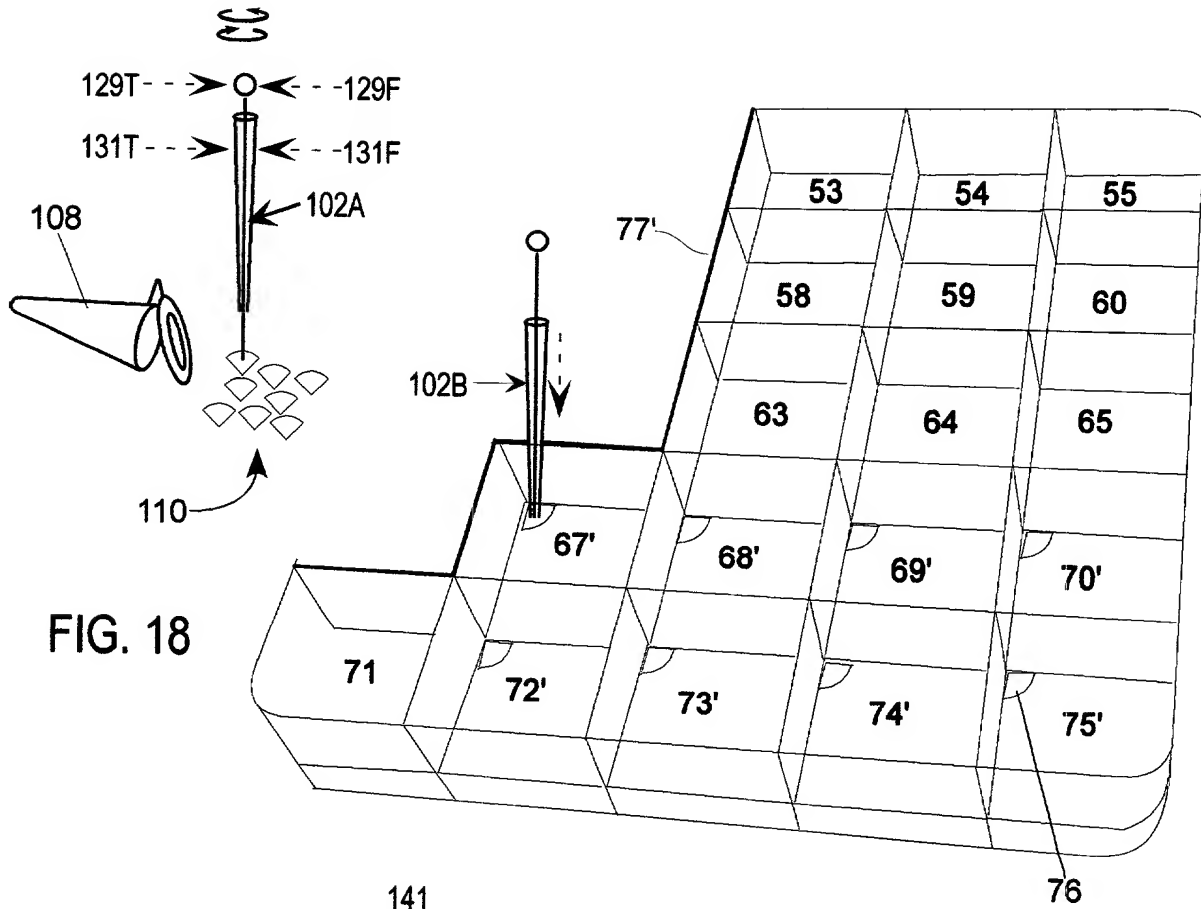


FIG. 18

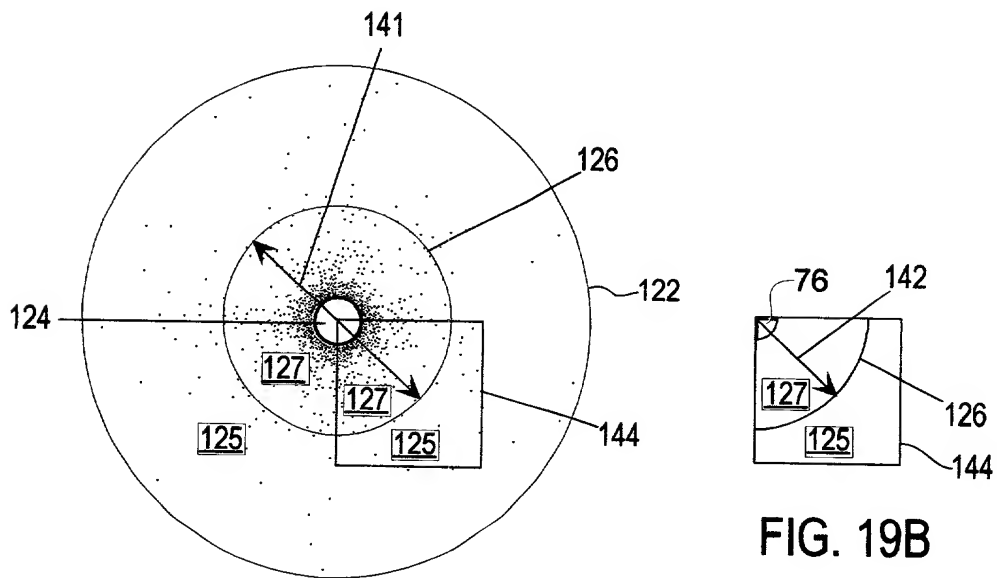


FIG. 19A

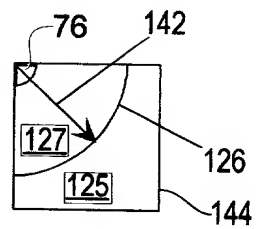


FIG. 19B



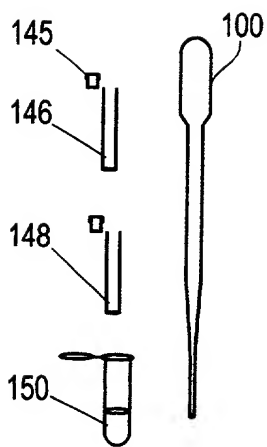


FIG. 20A

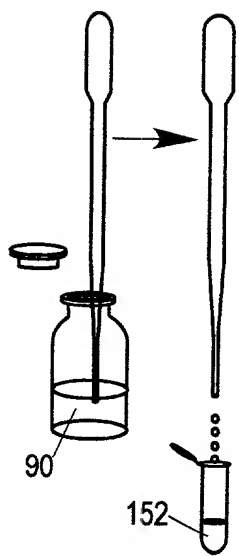


FIG. 20B



FIG. 20C

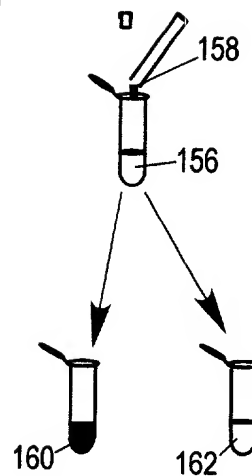


FIG. 20D

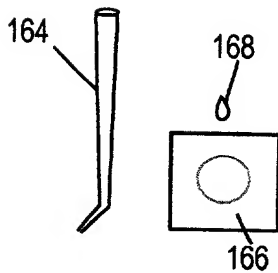


FIG. 21A

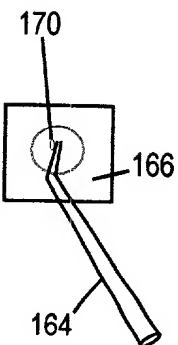


FIG. 21B

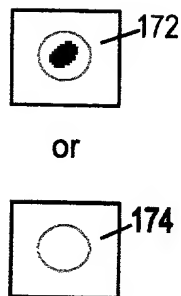


FIG. 21C

FIG. 20A



TABLE OF CULTURE PLATE MEDIA AND THEIR PURPOSES

51		Used for isolating a wide variety of microorganisms. Contains 5% sheep red blood cells. Will grow all non-fastidious gram negative and gram positive organisms.
56		Used for isolating, differentiating and presumptively identifying group D streptococcus and Enterococcus. These organisms cause the formation of a dark brown or black complex in the agar.
61	Mannitol salt agar	Staphylococci will grow in this medium while the growth of most other bacteria will be inhibited.
66		This medium permits the isolation of coagulase positive staphylococcus. Coagulase negative staph. and other bacteria are completely inhibited. Coagulase positive staph. reduce tellurite and produce black colonies.
71		Used for isolation of fungi. Suitable for growth of pathogenic fungi. Incubate for several days. Molds and yeasts form non-spreading, discrete colonies.
52		Used in the isolation of gram positive organisms from clinical and non-clinical specimens. Azide suppresses the growth of gram negative bacteria. Useful in determining hemolytic reactions.
62		Gram negative organisms that are able to metabolize citrate will grow luxuriantly. The medium is alkalized and changes from its initial green to deep blue.
63	Pseudomonas agar F	Used for differentiating Pseudomonas aeruginosa from other pseudomonads based on fluorescein production. Visible with UV lamp at 365nm.
64	Pseudomonas agar P	Used for differentiating Pseudomonas aeruginosa from other pseudomonads based on the production of pyocyanin, a non-fluorescent blue pigment.
60		Used to isolate and differentiate Salmonella. Colonies are greenish blue, with black centers
65		MacConkey agar with an added substrate(MUG) that becomes fluorescent when E.coli is present. The E.coli's beta-glucuronidase enzyme cleaves the colorless MUG to fluorescent product that is detected with UV light 365nm
53		MacConkey agar with lactose. Selective and differential medium for growing gram negative bacilli. Lactose fermenting strains grow as red or pink colonies.
54		MacConkey agar with glucose. Selective and differential medium for growing gram negative bacilli. Glucose fermenting strains grow as red or pink colonies.
55		MacConkey agar with mannitol. Selective and differential medium for growing gram negative bacilli. Mannitol fermenting strains grow as red or pink colonies.
57		MacConkey agar with inositol. Selective and differential medium for growing gram negative bacilli. Inositol fermenting strains grow as red or pink colonies.
58		MacConkey agar with Sucrose. Selective and differential medium for growing gram negative bacilli. Sucrose fermenting strains grow as red or pink colonies.
59		MacConkey agar with arabinose. Selective and differential medium for growing gram negative bacilli. Arabinose fermenting strains grow as red- pink colonies.
67-70 72-75	Mueller Hinton agar	Considered to be the best media for routine susceptibility testing of non-fastidious bacteria. Eight chambers are set aside for this purpose.

FIG. 22

FIG. 23

TABLE OF PREFERRED EMBODIMENT OF CULTURE PLATE MEDIA

Mannitol salt agar		Pseudomonas agar F	Pseudomonas agar P	
	Mueller Hinton agar	Mueller Hinton agar	Mueller Hinton agar	Mueller Hinton agar
	Mueller Hinton agar	Mueller Hinton agar	Mueller Hinton agar	Mueller Hinton agar
51	52	53	54	55
56	57	58	59	60
61	62	63	64	65
66	67	68	69	70
71	72	73	74	75

Fig. 24A

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TABLE FOR IDENTIFICATION OF NON-FASTIDIOUS GRAM NEGATIVE BACTERIA USING KIT RESULTS												
PAGE 1 OF 3												
IDENTIFICATION CRITERIA:	CIT	ARA	GLU	INO	LAC	MAN	SUC	OXI	NO2	MOT	MAC	
OBSERVE EACH CHAMBER OF PLATE REFERRING TO THE "TABLE OF CULTURE PLATE MEDIA AND THEIR PURPOSES". WHEN THE ORGANISM IS GRAM NEGATIVE (I.E. GROWTH ON MAC AND/OR OXIDASE POSTIVE) A LIST OF IDENTIFICATION CRITERIA CAN BE ASSEMBLED: CIT=UTILIZES CITRATE? ARA=FERMENTS ARABINOSE?; GLU=FERMENTS GLUCOSE?; INO=FERMENTS INOSITOL?; LAC=FERMENTS LACTOSE?; MAN=FERMENTS MANNITOL?; SUC=FERMENTS SUCROSE?; MAC=GROWS ON ANY MAC MEDIA; SEE ACCESSORY RESULTS FOR OXI=OXIDASE ACTIVITY, NO2=NITRATE REDUCTASE ACTIVITY,AND MOT=IS BACTERIA MOTILE?												
LET 1 = YES AND 0 = NO. FILL IN BOXES ACCORDINGLY AND THEN SEARCH DATABASE BELOW FOR BEST MATCH (MANUALLY OR USING DATABASE FILTER SOFTWARE)												

GRAM NEGATIVE ORGANISM	INCUBATION	CIT	ARA	GLU	INO	LAC	MAN	SUC	OXI	NO2	MOT	MAC
GRAM NEGATIVE ORGANISM	INCUBATION	CIT	ARA	GLU	INO	LAC	MAN	SUC	OXI	NO2	MOT	MAC
<i>Cedecea davisae</i>	12-20h	0	0	1	0	0	1	1	0	1	1	1
<i>Cedecea lapagei</i>	12-20h	1	0	1	0	0	1	0	0	1	1	1
<i>Cedecea neteri</i>	12-20h	1	0	1	0	0	1	1	0	1	1	1
<i>Cedecea sp. 3</i>	12-20h	1	0	1	0	0	1	1	0	1	1	1
<i>Cedecea sp.5</i>	12-20h	0	0	1	0	0	1	1	0	1	1	1
<i>Citrobacter amalonaticus</i>	12-20h	1	1	1	0	1	1	0	0	1	1	1
<i>Citrobacter diversus</i>	12-20h	1	1	1	0	1	1	0	0	1	1	1
<i>Citrobacter freundii</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Edwardsiella tarda</i>	12-20h	0	0	1	0	0	0	0	0	1	1	1
<i>Enterobacter aerogenes</i>	12-20h	1	1	1	1	1	1	1	0	1	1	1
<i>Enterobacter agglomerans</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Enterobacter amnigenus 1</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Enterobacter amnigenus 2</i>	12-20h	1	1	1	0	1	1	0	0	1	1	1
<i>Enterobacter cloacae</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Enterobacter gergoviae</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Enterobacter intermedium</i>	12-20h	1	1	1	0	1	1	0	0	1	1	1
<i>Enterobacter sakazakii</i>	12-20h	1	1	1	1	1	1	1	0	1	1	1
<i>Enterobacter taylorae</i>	12-20h	1	1	1	0	1	1	0	0	1	1	1
<i>Escherichia coli</i>	12-20h	0	1	1	0	1	1	0	0	1	1	1
<i>Escherichia fergusonii</i>	12-20h	0	1	1	0	1	1	0	0	1	1	1
<i>Escherichia hermannii</i>	12-20h	0	1	1	0	1	1	1	0	1	1	1
<i>Escherichia vulneris</i>	12-20h	0	1	1	0	1	1	0	0	1	1	1
<i>Ewingella americana</i>	12-20h	0	0	1	0	0	1	0	0	1	1	1
<i>Hafnia alvei</i>	12-20h	0	1	1	0	0	1	0	0	1	1	1
<i>Klebsiella oxytoca</i>	12-20h	1	1	1	1	1	1	1	0	1	0	1
<i>Klebsiella ozaenae</i>	12-20h	0	1	1	1	1	1	0	0	1	0	1
<i>Klebsiella pneumoniae</i>	12-20h	1	1	1	1	1	1	1	0	1	0	1
<i>Klebsiella Rhinoscleromatis</i>	12-20h	0	0	1	1	1	1	0	0	1	0	1
<i>Kluyvera sp.</i>	12-20h	1	1	1	0	0	1	1	0	1	1	1
<i>Moellerella wisconsensis</i>	12-20h	0	0	1	0	0	0	1	0	1	0	1
<i>Morganella morganii</i>	12-20h	0	0	1	0	0	0	0	0	1	1	1
<i>Presumptive Yersinia pestis</i>	12-20h	0	0	1	0	0	1	0	0	1	0	1
<i>Proteus mirabilis</i>	12-20h	1	0	1	0	0	0	0	0	1	1	1

Fig. 24B

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TABLE FOR IDENTIFICATION OF NON-FASTIDIOUS GRAM NEGATIVE BACTERIA USING KIT RESULTS												
PAGE 2 OF 3												
GRAM NEGATIVE ORGANISM	INCUBATION	CIT	ARA	GLU	INO	LAC	MAN	SUC	OXI	NO2	MOT	MAC
<i>Proteus penneri</i>	12-20h	0	0	1	0	0	0	1	0	1	1	1
<i>Proteus vulgaris</i>	12-20h	0	0	1	0	0	0	1	0	1	1	1
<i>Providencia alcalifaciens</i>	12-20h	1	0	1	0	0	0	0	0	1	1	1
<i>Providencia rettgeri</i>	12-20h	1	0	1	1	0	1	0	0	1	1	1
<i>Providencia stuartii</i> Ure-	12-20h	1	0	1	1	0	0	0	0	1	1	1
<i>Providencia stuartii</i> Ure+	12-20h	1	0	1	1	0	0	1	0	1	1	1
<i>Salmonella cholerae</i> suis	12-20h	0	0	1	0	0	1	0	0	1	1	1
<i>Salmonella enteritidis</i>	12-20h	1	1	1	0	0	1	0	0	1	1	1
<i>Salmonella paratyphi</i> A	12-20h	0	1	1	0	0	1	0	0	1	1	1
<i>Salmonella</i> subgroup 3	12-20h	1	1	1	0	0	1	0	0	1	1	1
<i>Salmonella typhi</i>	12-20h	0	0	1	0	0	1	0	0	1	1	1
<i>Salmonella typhimurium</i>	12-20h	1	1	1	0	0	1	0	0	1	1	1
<i>Serratia fonticola</i>	12-20h	0	1	1	1	1	1	0	0	1	1	1
<i>Serratia odorifera</i> 1	12-20h	1	1	1	1	1	1	1	0	1	1	1
<i>Serratia odorifera</i> 2	12-20h	1	1	1	1	1	1	0	0	1	1	1
<i>Serratia plymuthica</i>	12-20h	0	1	1	1	1	1	1	0	1	1	1
<i>Serratia rubidaea</i>	12-20h	1	1	1	0	1	1	1	0	1	1	1
<i>Serratia liquefaciens</i>	12-20h	1	1	1	1	1	1	1	0	1	1	1
<i>Serratia marcescens</i>	12-20h	1	0	1	1	1	1	1	0	1	1	1
<i>Tatumella ptyseos</i>	12-20h	0	0	1	0	0	0	1	0	1	0	1
<i>Yersinia enterocolitica</i>	12-20h	0	1	1	0	0	1	1	0	1	0	1
<i>Yersinia frederiksenii</i> /intermedia	12-20h	0	1	1	0	0	1	1	0	1	0	1
<i>Yersinia kristensenii</i>	12-20h	0	1	1	1	0	1	0	0	1	0	1
<i>Yersinia pseudotuberculosis</i>	12-20h	0	0	1	0	0	1	0	0	1	0	1
<i>Yersinia ruckeri</i> (AN)	12-20h	0	0	1	0	0	1	0	0	1	0	1
<i>Achromobacter</i> spp. (Vd)	24h	0	0	0	0	0	0	0	1	1	1	1
<i>Achromobacter</i> spp. (Vd)	48h	1	0	0	0	0	0	0	1	1	1	1
<i>Achromobacter xylosoxidans</i>	24h	0	0	0	0	0	0	0	1	1	1	1
<i>Achromobacter xylosoxidans</i>	48h	1	0	0	0	0	0	0	1	1	1	1
<i>Acinetobacter calcoaceticus</i> v. lwoffii	24h	0	0	0	0	0	0	0	0	0	0	1
<i>Acinetobacter calcoaceticus</i> v. lwoffii	48h	0	0	0	0	0	0	0	0	0	0	1
<i>Acinetobacter calcoaceticus</i> v. anitratus	24h	0	1	1	0	0	0	0	0	0	0	1
<i>Acinetobacter calcoaceticus</i> v. anitratus	48h	1	1	1	0	0	0	0	0	0	0	1
<i>Aeromonas hydrophila</i> group	24h	0	1	1	0	0	1	1	1	1	1	1
<i>Aeromonas salmonicida</i> (25c)	24h	0	0	1	0	0	1	0	1	1	0	1
<i>Agrobacterium radiobacter</i>	24h	0	0	0	0	0	0	0	1	0	1	1
<i>Agrobacterium radiobacter</i>	48h	1	0	0	0	0	0	0	1	0	1	1
<i>Alcaligenes</i> spp.	24h	0	0	0	0	0	0	0	1	0	1	1
<i>Alcaligenes</i> spp.	48h	1	0	0	0	0	0	0	1	0	1	1
<i>Bordetella bronchiseptica</i>	24h	0	0	0	0	0	0	0	1	1	1	1
<i>Bordetella bronchiseptica</i>	48h	1	0	0	0	0	0	0	1	1	1	1
CDC Group II J	24h	0	0	0	0	0	0	0	1	0	0	0
CDC Group II J	48h	0	0	0	0	0	0	0	1	0	0	0
CDC Group IV C-2	24h	0	0	0	0	0	0	0	1	0	1	1
CDC Group IV C-2	48h	1	0	0	0	0	0	0	1	0	1	1
CDC Group IV E	24h	0	0	0	0	0	0	0	1	1	0	1
CDC Group IV E	48h	0	0	0	0	0	0	0	1	1	0	1
CDC Group V E-1	24h	1	1	1	0	0	0	0	0	0	1	1
CDC Group V E-1	48h	1	1	1	0	0	0	0	0	1	1	1

TABLE FOR IDENTIFICATION OF NON-FASTIDIOUS GRAM NEGATIVE BACTERIA USING KIT RESULTS												
PAGE 3 OF 3												
GRAM NEGATIVE ORGANISM	INCUBATION	CIT	ARA	GLU	INO	LAC	MAN	SUC	OXI	NO2	MOT	MAC
CDC Group V E-2	24h	1	1	1	0	0	0	0	0	0	1	1
CDC Group V E-2	48h	1	1	1	0	0	0	0	0	0	1	1
CDS Group II F	24h	0	0	0	0	0	0	0	1	0	0	0
CDS Group II F	48h	1	0	0	0	0	0	0	1	0	0	0
Chromobacterium	24h	1	0	1	0	0	0	0	1	1	1	1
Chromobacterium	48h	1	0	1	0	0	0	0	1	1	1	1
Eikenella corrodens	24h	0	0	0	0	0	0	0	1	0	0	0
Eikenella corrodens	48h	0	0	0	0	0	0	0	1	0	0	0
Flavobacterium breve	24h	0	0	0	0	0	0	0	1	0	0	1
Flavobacterium breve	48h	1	0	0	0	0	0	0	1	0	0	1
Flavobacterium meningosepticum	24h	0	0	0	0	0	0	0	1	0	0	1
Flavobacterium meningosepticum	48h	1	0	0	0	0	0	0	1	0	0	1
Flavobacterium Multivorum	24h	0	0	1	0	0	0	0	1	0	0	1
Flavobacterium Multivorum	48h	1	0	1	0	0	0	1	1	0	0	1
Flavobacterium odoratum	24h	0	0	0	0	0	0	0	1	0	0	1
Flavobacterium odoratum	48h	1	0	0	0	0	0	0	1	0	0	1
Flavobacterium spiritivorum	24h	0	0	0	0	0	0	0	1	0	0	0
Flavobacterium spiritivorum	48h	0	0	0	0	0	0	0	1	0	0	0
Flavobacterium spp. (IIB)	24h	0	0	0	0	0	0	0	1	0	0	1
Flavobacterium spp. (IIB)	48h	1	0	0	0	0	0	0	1	0	0	1
Moraxella spp.	24h	0	0	0	0	0	0	0	1	1	0	0
Moraxella spp.	48h	0	0	0	0	0	0	0	1	1	0	0
Other Pseudomonas spp.	24h	0	0	0	0	0	0	0	1	1	1	1
Other Pseudomonas spp.	48h	1	0	0	0	0	0	0	1	1	1	1
Pasteurella aerogenes	24h	0	1	1	1	0	0	1	1	1	0	1
Pasteurella aerogenes	48h	0	1	1	1	0	0	1	1	1	0	1
Pasteurella multocida	24h	0							1	1	0	0
Pasteurella multocida	48h	0							1	1	0	0
Pasteurella-Actinobacillus spp.	24h	0	0	0	0	0	0	0	1	1	0	0
Pasteurella-Actinobacillus spp.	48h	0	0	0	0	0	0	0	1	1	0	0
Plesiomonas shigelloides	24h	0	0	1	1	0	0	0	1	1	1	1
Pseudomonas cepacia	24h	1	0	1	0	0	0	0	1	0	1	1
Pseudomonas cepacia	48h	1	0	1	0	0	0	0	1	0	1	1
Pseudomonas fluorescens	24h	0	0	0	0	0	0	0	1	1	1	1
Pseudomonas fluorescens	48h	1	0	0	0	0	0	0	1	1	1	1
Pseudomonas maltophilia	24h	1	0	0	0	0	0	0	0	0	1	1
Pseudomonas maltophilia	48h	1	0	0	0	0	0	0	0	0	1	1
Pseudomonas Paucimobilis	24h	0	0	0	0	0	0	0	1	0	0	0
Pseudomonas Paucimobilis	48h	1	0	0	0	0	0	0	1	0	0	0
Pseudomonas putida	24h	1	0	0	0	0	0	0	1	0	1	1
Pseudomonas putida	48h	1	0	0	0	0	0	0	1	0	1	1
Pseudomonas putrefaciens	24h	1	0	0	0	0	0	0	1	1	1	1
Pseudomonas putrefaciens	48h	1	0	0	0	0	0	0	1	1	1	1
Pseudomonas Stutzeri	24h	0	0	0	0	0	0	0	1	1	1	1
Pseudomonas Stutzeri	48h	1	0	0	0	0	0	0	1	1	1	1
Pseudomonas aeruginosa	24h	1	0	1	0	0	0	0	1	1	1	1
Pseudomonas aeruginosa	48h	1	0	1	0	0	0	0	1	1	1	1
Pseudomonas pseudomallei	24h	0	1	0	1	0	1	1	1	1	1	1
Pseudomonas pseudomallei	48h	0	0	1	1	0	1	1	1	1	1	1

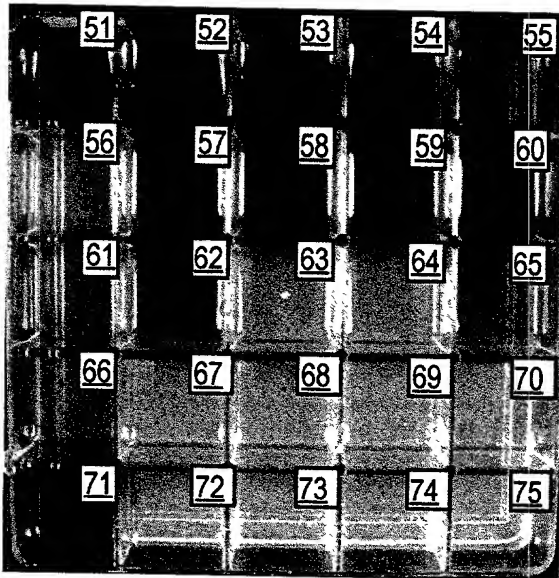


FIG. 25

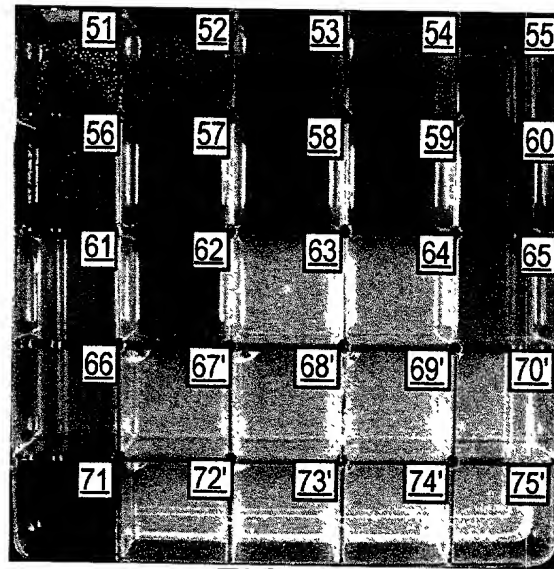


FIG. 26

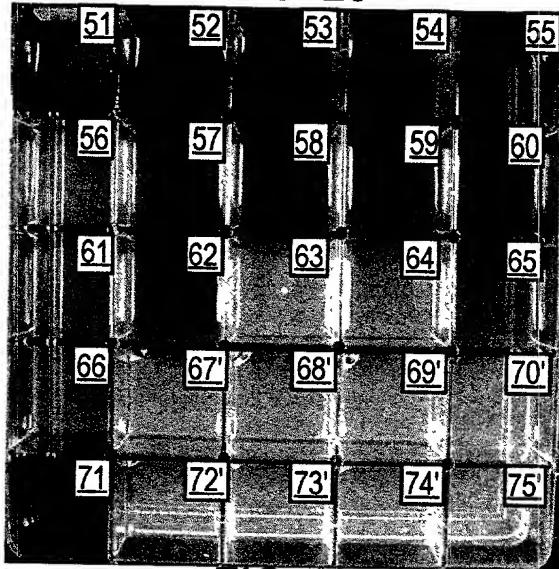


FIG. 27

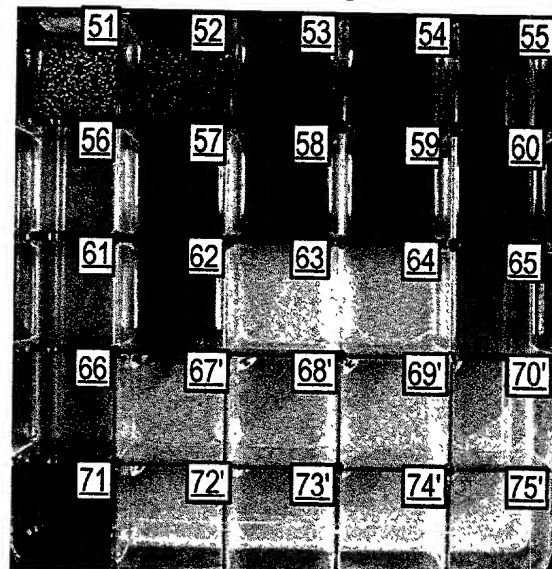


FIG. 28

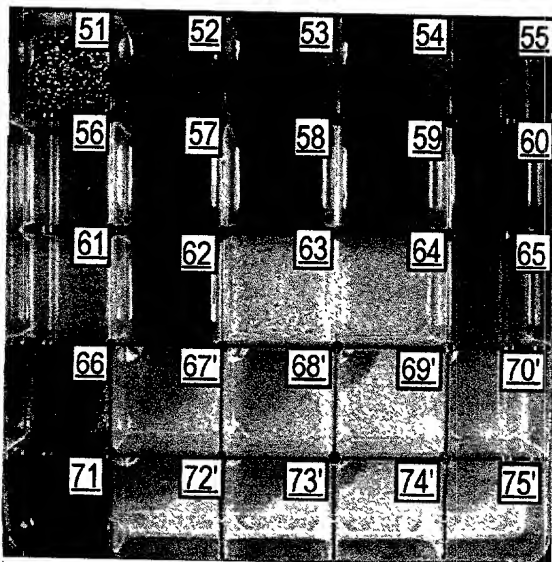


FIG. 29

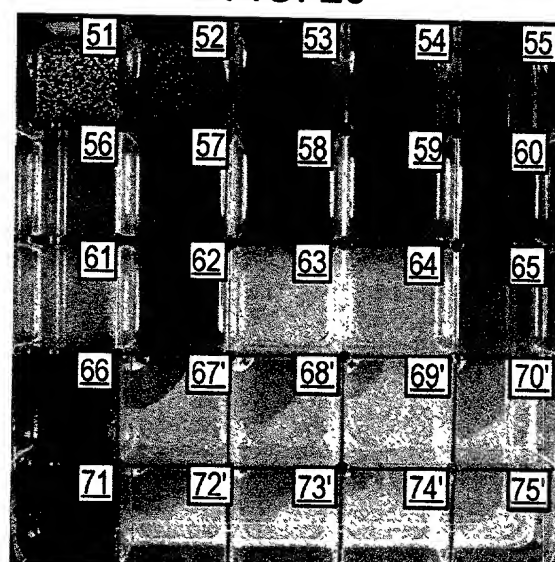


FIG. 30

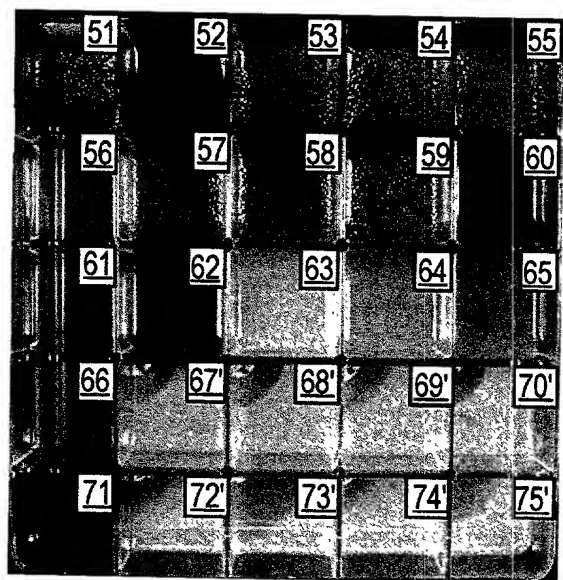


FIG. 31

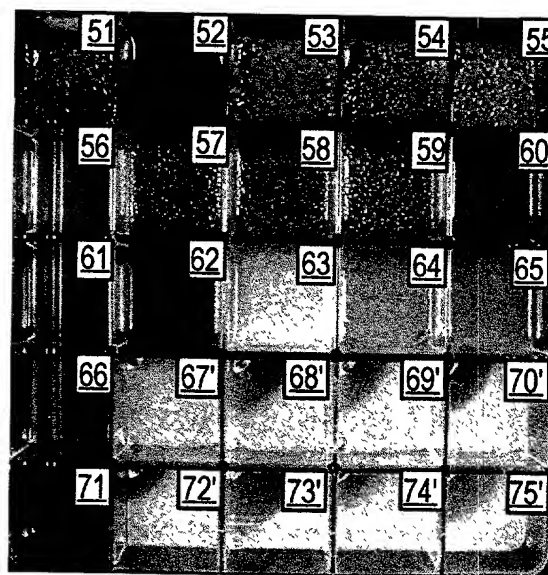


FIG. 32

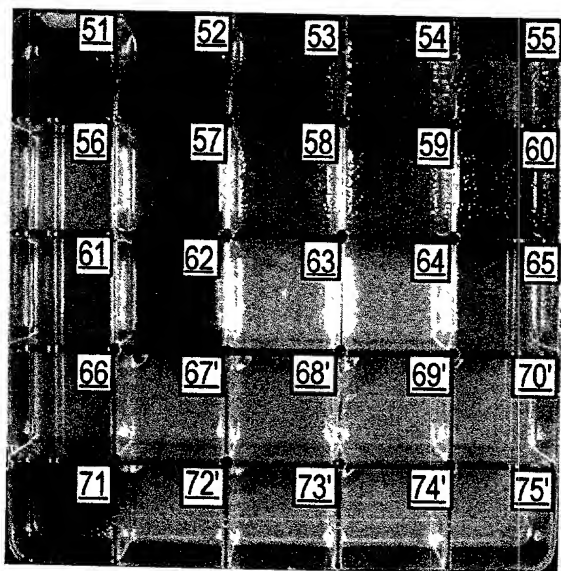


FIG. 33

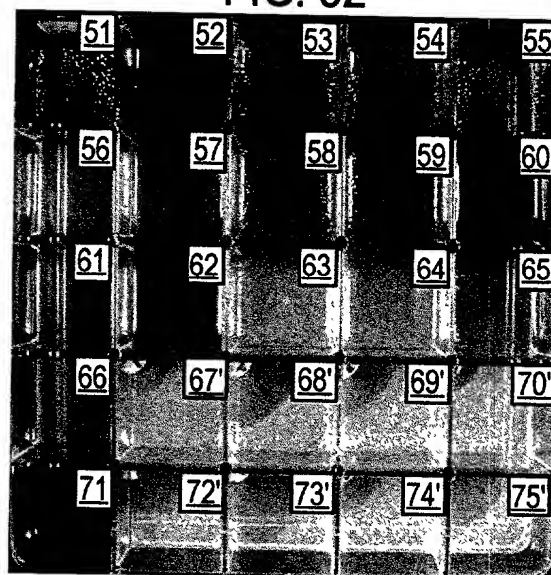


FIG. 34

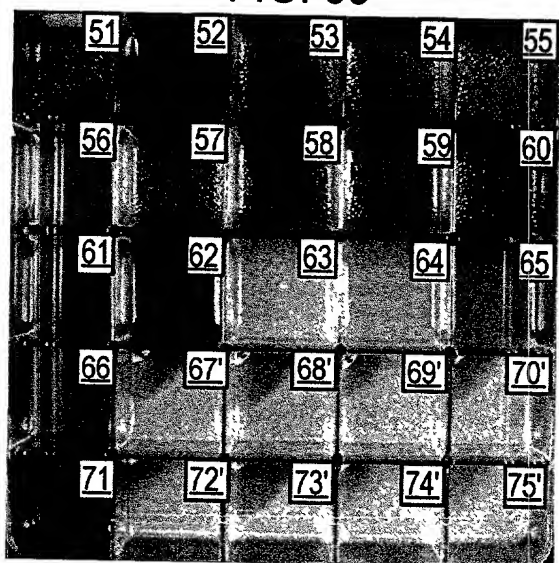


FIG. 35

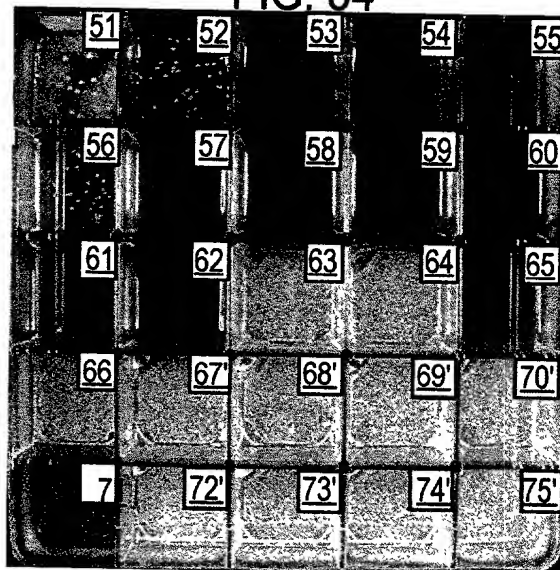
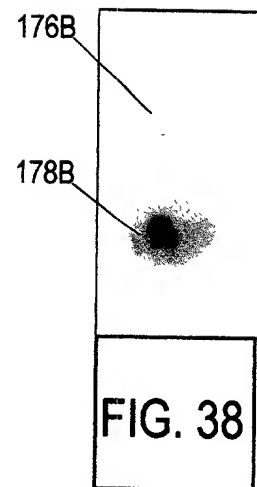
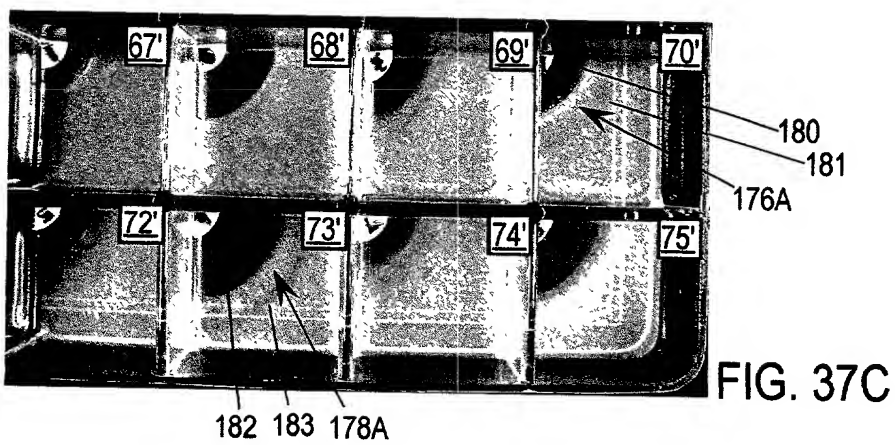
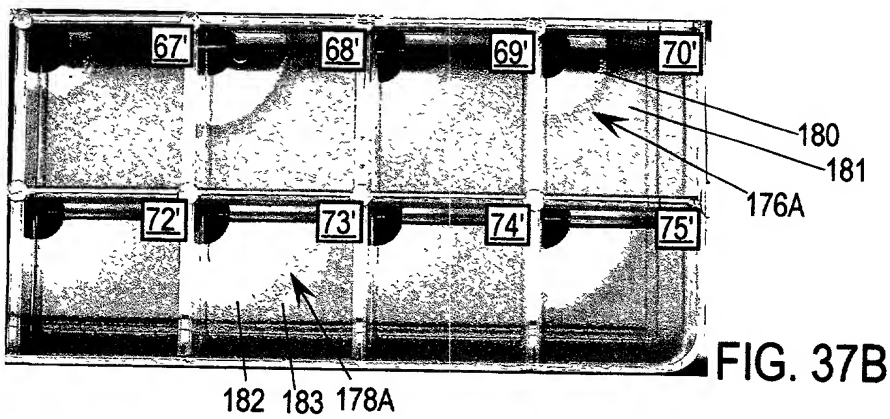
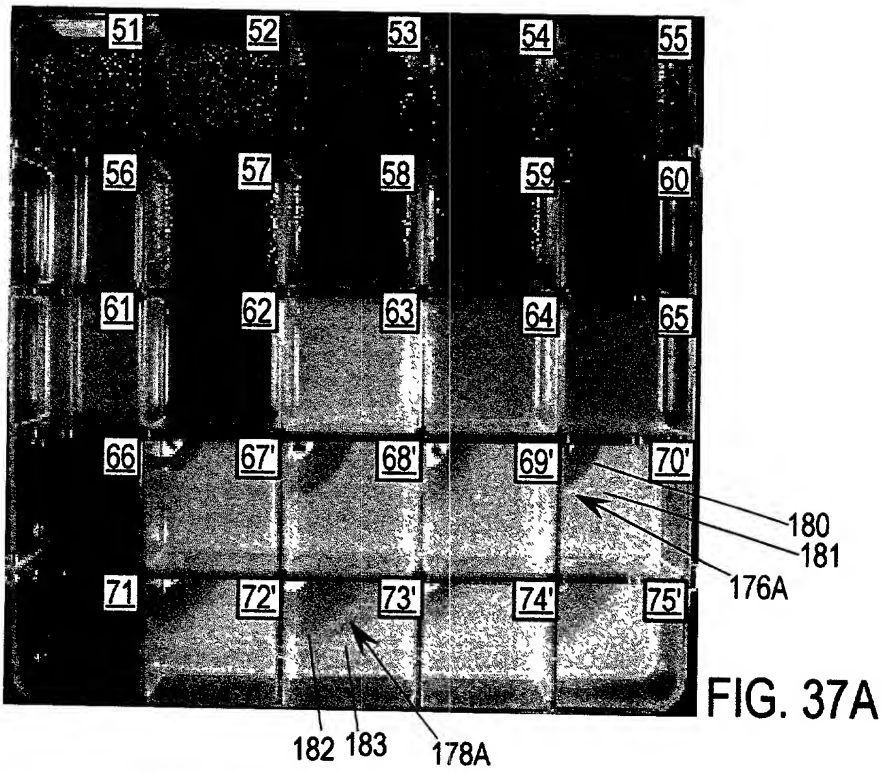


FIG. 36



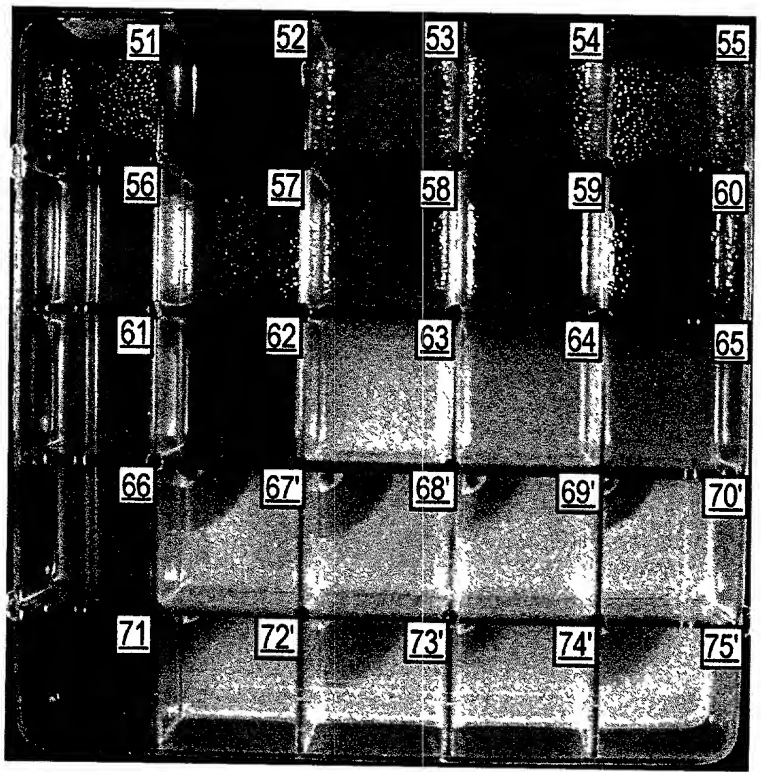


FIG. 39A

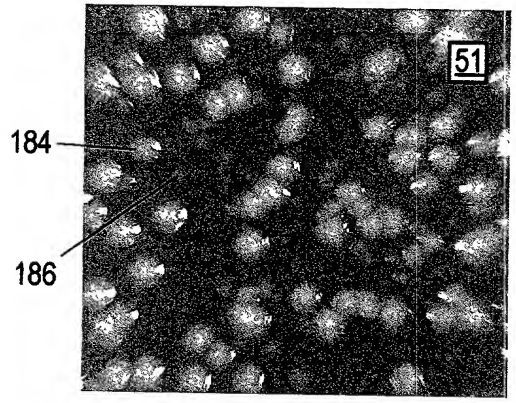


FIG. 39B

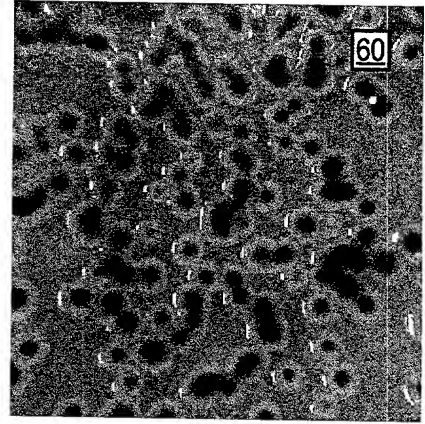


FIG. 39C



FIG. 39D

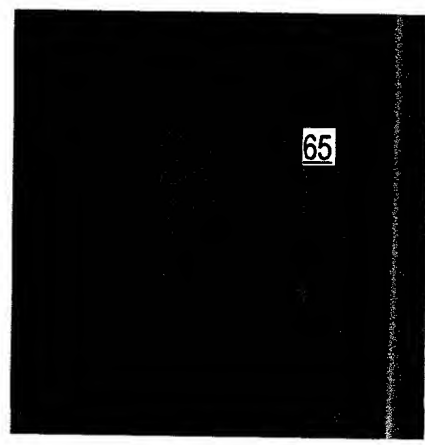


FIG. 39E

Inventor's note:

This page at one time contained figures. They have recently been deleted and are not part of my final application. Rather than having to reprint all of my drawings, I have included this page as a blank page.

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